Insights into the Electronic Structure of Cu^{II} Bound to an Imidazole Analogue of Westiellamide

Peter Comba,[†] Nina Dovalil,^{†,‡} Graeme R. Hanson,^{*,‡} Jeffrey R. Harmer,[‡] Christopher J. Noble,[‡] Mark J. Riley,[§] and Bjoern Seibold[†]

[†]Anorganisch-Chemisches Institut, Universität Heidelberg, D-69120 Heidelberg, Germany

[‡]Centre for Advanced Imaging Brisbane and [§]School of Chemistry and Molecular Biosciences, The University of Queensland, Queensland 4072, Australia

Supporting Information

ABSTRACT: Three synthetic analogues of westiallamide, H_3L^{wa} , have previously been synthesized (H_3L^{1-3}) that have a common backbone (derived from L-valine) with H_3L^{wa} but differ in their heterocyclic rings (imidazole, oxazole, thiazole, and oxazoline). Herein we explore in detail through highresolution pulsed electron paramagnetic resonance (EPR) and magnetic circular dichroism (MCD) spectroscopy in conjunction with density functional theory (DFT) the geometric and electronic structures of the mono- and dinuclear Cu^{II}



complexes of these cyclic pseudo hexapeptides. Orientation-selective hyperfine sublevel correlation, electron nuclear double resonance, and three-pulse electron spin echo envelope modulation spectroscopy of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ reveal delocalization of the unpaired electron spin onto the ligating and distal nitrogens of the coordinated heterocyclic rings and that they are magnetically inequivalent. DFT calculations confirm this and show similar spin densities on the distal heteroatoms in the heterocyclic rings coordinated to the Cu^{II} ion in the other cyclic pseudo hexapeptide $[Cu^{II}(H_2L^{2,3,wa})(MeOH)_2]^+$ complexes. The magnetic inequivalencies in $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ arise from different orientations of the heterocyclic rings coordinated to the Cu^{II} ion, and the delocalization of the unpaired electron onto the distal heteroatoms within these Nmethylimidazole rings depends upon their location with respect to the Cu^{II} $d_{x^2-y^2}$ orbital. A systematic study of DFT functionals and basis sets was undertaken to examine the ability to reproduce the experimentally determined spin Hamiltonian parameters. Inclusion of spin-orbit coupling (SOC) using MAG-ReSpect or ORCA with a BHLYP/IGLO-II Wachters setup with SOC corrections and ~38% Hartree-Fock exchange gave the best predictions of the g and $A(^{63}Cu)$ matrices. DFT calculations of the ¹⁴N hyperfine and quadrupole parameters for the distal nitrogens of the coordinated heterocyclic rings in $[Cu^{II}(H_2L^1) (MeOH)_2$ ⁺ with the B1LYP functional and the SVP basis set were in excellent agreement with the experimental data, though other choices of functional and basis set also provided reasonable values. MCD, EPR, mass spectrometry, and DFT showed that preparation of the dinuclear Cu^{II} complex in a 1:1 MeOH/glycerol mixture (necessary for MCD) resulted in the exchange of the bridging methoxide ligand for glycerol with a corresponding decrease in the magnitude of the exchange coupling.

INTRODUCTION

Ascidiacea (ascidians or sea squirts) are sessile filter-feeding marine invertebrate animals that are normally found firmly attached to a substratum such as rocks along coastlines or coral reefs, where the water levels are shallow and the salinity is greater than 2.5%.¹⁻⁶ They have a sac-like body that incorporates three regions, the pharyngeal, abdomen, and postabdomen that is housed within a tough outer "tunic" made of the polysaccharide tunicin, which compared to other tunicates leads to a more rigid "exoskeleton".¹⁻⁶ The pharyngeal region contains the pharynx, which is the basis of the digestive system involving filtering plankton, metal ions, and nutrients out of the seawater through its two siphons. The abdomen contains the heart and gonads.¹⁻⁶ Ascidians can be found all over the world and are among those marine organisms whose Cu^{II} accumulation is high.^{7,8} The fact that

 ${\rm Cu}^{\rm II}$ is found in nonpolar tissue fractions suggests its complexation by low molecular weight organic molecules, such as natural cyclic pseudo peptides, for example, patellamides or proteins. Indeed, the marine genus *Lissoclinum* is a rich source of cyclic peptide alkaloids, featuring multiple oxazoline, thiazoline, oxazole, or thiazole rings that have interesting cytotoxicity, antibacterial, and antiviral properties.^{8–11} Ascidians live in a symbiotic relationship with prochloron, a unicellular oxygenic photosynthetic prokaryote belonging to the cyanobacteria phylum, and it is thought that it is responsible for the synthesis of these peptides.^{12–16} The Cu^{II} coordination chemistry of the native cyclic pseudo octa- and hexapeptides purified from *L. patella* and *L. bistratum*, respectively, and chemically synthesized model cyclic pseudo

Received: June 19, 2014 **Published:** November 13, 2014 peptides for these two classes has been extensively studied.^{7,11,17–28} While the biological function of these cyclic pseudo hexa- and octapeptides is at present unknown, the fact that there is a diverse range of these peptides and that many^{15,16} are synthesized at the ribosome in an archaic symbiont indicates that they have an important biological role. We have previously shown that dinuclear Cu^{II} complexes of cyclic pseudo octapeptides can catalyze CO₂ fixation (most efficient small molecular weight catalysts known to date) producing carbonate (the backbone of a coral reef),²⁷ and also mono- and diphosphoester hydrolysis.²⁸ Whether or not the corresponding Cu^{II} pseudo hexapeptide complexes have a metabolic role has to our knowledge not been reported in the literature.

Westiellamide, a cyclic pseudo hexapeptide $(H_3L^{wa}, Chart 1)$ that has been isolated from the marine genus *L. bistratum* and



from the terrestrial genus *Westiellopsis prolifica*, is known to accumulate in leukemia cells and is found to inhibit cytokinesis.²⁹ In solution, the metal-free macrocycle H_3L^{wa} adopts C_3 symmetry (X-ray crystal structures shown in Figure 1), where the heterocyclic and amide nitrogen atoms point toward the inside of the macrocycle, and the isopropyl residues face to the same side of the macrocycle plane.³⁰ The five-membered heterocyclic oxazoline rings result from condensation of threonine side chains with the preceding carbonyl

groups of the valine residues in the peptide sequence. Three synthetic analogues (Chart 1) of H_3L^{wa} have been prepared (H_3L^{1-3}) that have a common backbone with H_3L^{wa} and differ solely in their heterocyclic donor groups (*N*-methylimidazole, oxazole, and thiazole).³¹ Plots of their X-ray structures are shown in Figure 1.^{30,31} Importantly, the difference in the heterocyclic rings results in subtle differences in the shape of the four macrocycles as the single (σ) bonds of the oxazoline rings of westiellamide H_3L^{wa} increase the flexibility of the macrocycle.

Initial studies on the metal complexation of westiellamide were performed by Wipf et al., who reported an unusual Ag₄ complex formed with westiellamide.³² In this complex three of the four Ag⁺ ions (coordinated to the oxazoline nitrogens) are located in a pseudo trigonal-planar arrangement about a central Ag⁺ ion, and all four ions are sandwiched between two westiellamide macrocycles. The central Ag⁺ ion is coordinated in a distorted octahedral arrangement by the carbonyl oxygen atoms of the two westiellamide molecules. In addition to the interaction of H₃L^{wa} with Ag⁺, Wipf et al. also observed weak interactions between westiellamide and other metal ions such as Na⁺, Cu⁺, Fe³⁺, Hg²⁺, Au²⁺, and Zn^{2+.32}

Investigations of the interaction of Cu^{II} with H₃L^{wa} and a series of synthetic analogues H₃L¹⁻³ (Figure 1) revealed that after addition of base all cyclic pseudo hexapeptides readily form stable mono- and dinuclear Cu^{II} complexes.²² In the absence of base, Cu^{II} forms a 2:1 (H_3L^2/Cu^{II}) complex in which Cu^{II} is coordinated to the outside of the macrocycle through the carbonyl oxygen atoms of the peptide bonds,²² in a similar manner to the central Ag⁺ ion in the unusual [Ag₄- $(H_3L^{wa})_2$] complex reported by Wipf et al.³² Previous investigations of the Cu^{II} coordination chemistry of the cyclic pseudo hexapeptides H₃L¹⁻³ and models for the natural cyclic pseudo peptide westiellamide H₃L^{wa} revealed interesting differences in their geometric and electronic structures.² While all cyclic pseudo hexapeptides form mononuclear $\mathrm{Cu}^{\mathrm{II}}$ complexes in the presence of 1 equivalent of base, Cu^{II} was coordinated to a $N_{het} - N_{amide} - N_{het}$ binding site in $H_3 L^{1-3}$ and to a $N_{het}-N_{amide}-N_{het}-N_{het}$ binding site in H_3L^{wa} (N_{het} : nitrogen atom of the heterocyclic ring; N_{amide} : deprotonated amide nitrogen). The coordination sphere is completed with



Figure 1. X-ray structures^{30,31} of H_3L^1 , H_3L^2 , H_3L^3 , and H_3L^{wa} showing the top (upper) and side (lower) views. Atom colors: nitrogen - blue, oxygen - red, carbon - gray, sulfur-yellow. Hydrogen atoms and solvent molecules were omitted for clarity. Blue, red, and green heterocyclic rings correspond to *N*-methylimidazole, oxazole/oxazoline, and thiazole rings, respectively.

Table 1. Anisotropic Spin Hamiltonian Parameters for	r the Mononuclear Complexes [C	$[u^{II}(H_2L^1)(MeOH)_2]^+$,
$[Cu^{II}(H_2L^2)(MeOH)_2]^+$, $[Cu^{II}(H_2L^3)(MeOH)_2]^+$, and	$[\operatorname{Cu}^{\mathrm{II}}(\mathrm{H}_{2}\mathrm{L}^{\mathrm{wa}})(\mathrm{MeOH})_{2}]^{+22 a}$	

	$[Cu^{II}(H_2L^1)]^+$	$[Cu^{II}(H_2L^2)]^+$	$[Cu^{II}(H_2L^3)]^+$	$[Cu^{II}(H_2L^{wa})]^+$
g _x	2.088	2.083	2.082	2.083
<i>g</i> _y	2.051	2.034	2.037	2.051
g_z	2.278	2.279	2.263	2.267
$ A_x $ (⁶³ Cu)	17.0	17.3	15.7	14.0
$ A_{y} $ (⁶³ Cu)	15.4	17.2	19.9	16.2
$ A_z $ (⁶³ Cu)	153.4	153.0	150.0	175
$ A_x $ (¹⁴ N _{het})	14.5	15.7	14.3	12.4
$ A_{\gamma} $ (¹⁴ N _{het})	7.1	7.1	7.0	6.2
$ A_z $ (¹⁴ N _{het})	9.0	9.0	9.0	10.4
$ A_x ({}^{14}N_{amide}) (+ {}^{14}N_{het})^b$	13.2	13.4	11.5	16.5
$ A_{y} $ (¹⁴ N _{amide}) (+ ¹⁴ N _{het}) ^b	15.2	14.1	15.7	12.7
$ A_z ({}^{14}N_{amide}) (+ {}^{14}N_{het})^b$	9.5	9.5	9.5	13.4

^{*a*}The ⁶³Cu and ¹⁴N hyperfine values $(10^{-4} \text{ cm}^{-1})$ were determined from computer simulation of the CW EPR spectra assuming two magnetically equivalent nitrogen $({}^{14}N_{het})$ nuclei. The nuclear quadrupole interaction was ignored. ^{*b*}For $[Cu^{II}(H_2L^{wa})]^+$.

one or two axially coordinated solvent molecules. While the binding motif $N_{amide}-N_{het}-N_{amide}$ is also possible, the mononuclear Cu^{II} complexes exhibit a strong preference for the $N_{het}-N_{amide}-N_{het}$ binding motif, while in the dinuclear Cu^{II} complexes of H_3L^{1-3} the Cu^{II} ions bind to both binding sites and are bridged by either methanol or methoxide, which helps stabilize the dinuclear complex.²² Cu^{II} complexation requires deprotonation of the amide nitrogen(s), which is metal ion assisted and takes place at relatively low pH values. The protons that are released upon coordination of Cu^{II} acidify the solution, and thus addition of base is mandatory to achieve complete complexation.^{22,23} Since seawater is slightly basic (pH \approx 8), it is likely that metal ions are coordinated to these cyclic pseudo hexapeptides and that these complexes may be involved in metal ion transport or have metabolic roles.

Herein, we extend our previous continuous wave electron paramagnetic resonance (CW EPR) measurements on the mono- and dinuclear Cu^{II} complexes of H_3L^{wa} and the series of synthetic analogues H_3L^{1-3} by undertaking high-resolution pulsed EPR and magnetic circular dichroism (MCD) studies in conjunction with density functional theory (DFT) calculations to further characterize the geometric and electronic structure of the mono- and dinuclear Cu^{II} complexes of H_3L^{1-3} as structural analogues of westiellamide, H_3L^{wa} . A systematic study of the choice of basis set and functional in DFT calculations was also undertaken to determine their suitability for the calculation of the spin Hamiltonian parameters for the $[Cu^{II}(H_2L^{1-3,wa})-(MeOH)_2]^+$ complexes.

EXPERIMENTAL SECTION

Materials. Cyclic peptides $H_3L^{1-3,wa}$ were prepared according to published procedures.^{10,33} All materials obtained commercially were of reagent grade and used without further purification. Triethylamine was obtained from Wako Chemicals (Tokyo, Japan). Tetrabutylammonium perchlorate and methanol were purchased from Fluka and Aldrich, respectively.

Methods. X-band (ca. 9.5 GHz) CW and pulsed EPR spectra were recorded with a Bruker Biospin Elexsys E580 EPR spectrometer fitted with either a superhigh Q cavity (CW EPR) or an ER 4118X-MD5 flexline resonator (pulsed EPR). The X-band (~9.4 GHz) CW EPR spectra were recorded under the following conditions: 140 K, modulation frequency 100 kHz, modulation amplitude 0.05 mT, and microwave power 20 mW (10 dB). The magnetic field and microwave frequency were calibrated with a Bruker ER 036TM Teslameter and a Bruker microwave frequency counter, respectively. An Oxford

Instruments flow-through cryostat (CF935LT) in conjunction with an Oxford Instruments ITC503 variable-temperature controller provided temperatures of 1.5-50 K at the sample position in the cavity. Spectrometer tuning, signal averaging, and visualization were accomplished with Bruker's Xepr (version 2.4b.12) software.

Three-pulse electron spin echo envelope modulation (ESEEM) and hyperfine sublevel correlation (HYSCORE)^{34,35} experiments were recorded at 5 K and 9.67 GHz and employed the pulse sequences $\pi/2$ - τ - $\pi/2$ - t_1 - $\pi/2$ - τ -echo and $\pi/2$ - τ - $\pi/2$ - t_1 - π - t_2 - $\pi/2$ - τ -echo, respectively. The following parameters were used: microwave pulse lengths $t_{\pi/2} = 16$ ns, $t_{\pi} = 32$ ns, $\tau = 140$ ns, starting times $t_{1,0} = t_{2,0} =$ 400 ns, and time increments of $\Delta t = 40$ ns. For three-pulse ESEEM experiments 256 points were collected for each trace, and for HYSCORE a 256×256 data matrix was collected. In both three-pulse ESEEM and HYSCORE experiments a four-step phase cycle was used to remove unwanted echoes. All data were processed with MATLAB 2013A (8.1.0.604, The MathWorks, Inc.). The time traces were baseline-corrected with an exponential, apodized with a Gaussian window, and zero filled. After a one-dimensional (three-pulse ESEEM) or two-dimensional (HYSCORE) Fourier transformation, absolutevalue spectra were calculated. X-band Davies electron nuclear double resonance (ENDOR) spectra were recorded at 9.67 GHz at 5 K using the microwave pulse sequence $\pi - T - \pi/2 - \tau - \pi - \tau$ - echo with microwave pulses of lengths $t_{\pi/2} = 24$ ns and $t_{\pi} = 32$ ns, with $\tau = 400$ ns. During time $T = 9 \ \mu s$ a radio frequency (RF) pulse of 6 μs was applied using 100% gain of a 150 W Applied Engineering RF amplifier.

CW and pulsed EPR spectra of the mono- and dinuclear complexes were simulated with the XSophe–Sophe-XeprView^{36,37} (version 1.1.4), Molecular Sophe^{37,38} (version 2.3.1), and EasySpin³⁹ computer simulation software suites on a personal computer running the Mandriva Linux v2010.2 operating system. The ENDOR and HYSCORE data were simulated with the program EasySpin³⁹ using the functions salt and saffron. HYSCORE cross-peak positions (frequencies) were matched to the experimental positions by calculating cross-peak frequencies by diagonalization of a spin Hamiltonian (eq 1) with nuclear Zeeman, hyperfine, and nuclear quadrupole interactions (no intensity calculation).

High-resolution electrospray ionization mass spectrometry (ESI-MS) was performed with a 9.4 T Bruker ApexQe Qh-ICR hybrid instrument with an Apollo II MTP ion source in the positive-ion electrospray ionization (ESI) mode. Sample solutions in methanol/ glycerol (1:1) at concentrations of $10^{-4}-10^{-5}$ M were admitted to the ESI interface by means of a syringe pump at 5 mL min⁻¹ and sprayed at 4.5 kV with a desolvation gas flow of 2.0 L min⁻¹ at 25 °C and a nebulizer gas flow of 1.0 L min⁻¹. The ions were accumulated in the storage hexapole for 0.1–1.0 s and then transferred into the ICR cell. Trapping was achieved at a sidekick potential of –4.0 V and trapping potentials of roughly 1 V. The mass spectra were acquired in the broadband mode with 1 M data points. Typically, 16 transients were accumulated for one magnitude spectrum. The instrument was controlled by Bruker ApexControl 2.0.0.beta software, and data analysis was performed using the Bruker DataAnalysis 3.4 software.

Density functional theory (DFT) calculations were performed with Gaussian 03,⁴⁰ Gaussian 09,⁴¹ TURBOMOLE,⁴² ORCA,⁴³ and MAG-ReSpect.⁴⁴ Geometry optimizations were performed using the B3LYP⁴⁵⁻⁴⁷ hybrid functional in combination with the basis sets 6-31g*⁴⁷ (C, H N, O, S) and TZVP⁴⁸ (Cu). Frequency calculations were performed subsequent to the geometry optimization to confirm the structures were at a potential minimum on the potential energy surface. Using these optimized geometries, we examined a wide range of basis sets and functionals to explore the accuracy in reproducing the experimental spin Hamiltonian matrices for the $[Cu(H_2L^{1/2,3,wa})-(MeOH)_2]^+$ complexes.

Magnetic circular dichroism (MCD) spectra were recorded on an instrument based on a high-throughput/high-resolution Jobin Yvon 750s monochromator and an Oxford Instruments SpectroMag superconducting magnet equipped with an SM4 Cryostat using either a Blue (Hamamatsu R7459) PMT (UV) or Si APD (Vis) detector. The MCD spectra were recorded in a methanol/glycerol 1:1 mixture at various temperatures and magnetic fields.

RESULTS AND DISCUSSION

EPR Spectroscopy. CW and pulsed EPR experiments are commonly exploited to gain insights into the Cu^{II} ion geometry, ligating atoms, singly occupied molecular orbital (SOMO), and the extent of delocalization of the unpaired electron spin (i.e., spin density) onto the ligating atoms and atoms within the second coordination sphere.^{22,34,49} X-band CW EPR spectra have already been reported, and the orthorhombic spin Hamiltonian parameters (*g*, *A*(⁶³Cu), *A*(¹⁴N_{amide}), *A*(¹⁴N_{het})) determined through computer simulation of the first and second derivative EPR spectra are reproduced in Table 1 to allow readers to compare the DFT and EPR results and also the CW EPR and pulsed EPR data presented herein.²²

We extended these CW EPR studies by performing orientation-selective pulsed ENDOR and three-pulse ESEEM and HYSCORE experiments³⁴ to characterize the hyperfine and nuclear quadrupole couplings to the ligating and distal nitrogen atoms in the N-methylimidazole rings of $[Cu^{II}(H_2L^1)]$ - $(MeOH)_2$ ⁺. These techniques provide insights into the extent of delocalization of the unpaired electron onto the heterocyclic rings and peptide backbone. Davies ENDOR spectra (Figure 2c) show signals from strongly coupled ¹⁴N nuclei with nuclear (ENDOR) frequencies in the range of 15-26 MHz that could be simulated using a model comprised of three nitrogen nuclei, namely, $2 \times N_{het}$ and $1 \times N_{amide}$ ³⁹ These simulations are shown in Figure 2c, and the optimized ¹⁴N hyperfine and nuclear quadrupole parameters are listed in Table 2. The ¹⁴N hyperfine values determined from the ENDOR data are very similar to the values determined previously from the X-band CW EPR data,²² though in this case the nuclear quadrupole interaction was neglected. Utilizing the g and $A(^{63}Cu)$ matrices determined from the CW EPR spectrum and the $A({}^{14}N)$ and $P({}^{14}N)$ matrices determined from the ENDOR spectra (Table 2) provides for an accurate simulation of the CW EPR spectrum (Figure 2a,b), though the perpendicular region of the spectrum is very sensitive to the choice of g- and A-strain line width parameters.^{50–52} The ¹⁴N hyperfine and quadrupole parameters (Table 2) for the ligating nitrogen atoms (N_{het} and N_{amide}) are typical of Cu-His (imidazole) complexes^{49,53} and Cu^{II} complexes containing coordinated peptide nitrogens.53,54 The Davies ENDOR spectrum also reveals ¹H peaks centered around the proton Larmor frequency (Figure 2c, $B_0 = 335$ mT).





Figure 2. CW EPR and Davies ENDOR spectra of $[Cu^{II}(H_2L^1)-(MeOH)_2]^+$ in methanol. (a) Second derivative X-band CW EPR spectrum (red) and (b) the corresponding simulation optimized to fit the pulsed and CW EPR data (blue). (c) X-band ($\nu = 9.672$ GHz) Davies ENDOR spectra (black) recorded at 5.0 K at the indicated field positions along with the simulations (total blue) for three strongly coupled ¹⁴N nuclei (green, red, and magenta). Spin Hamiltonian parameters are given in Table 2. The vertical lines show the ¹H Larmor frequency, and these proton resonances were not simulated.

The ¹H couplings (~4 MHz) were not included in the simulated spectra and arise from protons within ca. 3.4 Å (assuming dipole–dipole coupling). These may be assigned to either the backbone CH or equatorially coordinated MeOH ligands (CH₃ or OH if not deprotonated) moieties. Without additional ²H exchange data, a unique assignment cannot be made.

Smaller hyperfine couplings to distal nitrogens in the coordinated imidazole rings of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ were characterized with orientation-selective three-pulse ESEEM and HYSCORE experiments. Surface and contour plots of the HYSCORE spectrum (Figure 3a,b) recorded at the echo maximum of the EPR spectrum (near the g_x/g_y positions) allow the assignment of ¹⁴N single-quantum (s) and double-quantum (d) cross-peaks and initial guesses for the hyperfine and nuclear quadrupole couplings. Subscripted atom numbers correspond to those from the DFT calculations (see Figure 5). The experimental HYSCORE (Figure 3a,b) and orientationselective three-pulse ESSEM (Figure 3c-black) spectra clearly show a doubling of the single and double quantum resonances (see N-14a and N-14b, Figure 3b), indicating ¹⁴N hyperfine coupling to two different nitrogen nuclei. In addition the HYSCORE spectra (Figure 3a,b) clearly show cross-peaks (label N-34, Figure 3b) from a weakly coupled ¹⁴N nucleus.

				nucleus		
parameter	N _{het}	\mathbf{N}_{het}	N _{amide}	N _{distal} -14a	N _{distal} -14b	N _{distal} -34
$ A_x $ (¹⁴ N)	38.0	39.0	44.0	2.5	2.9	1.0
$ A_y $ (¹⁴ N)	37.0	37.5	35.0	2.5	2.9	1.0
$ A_z $ (¹⁴ N)	37.5	38.5	38.5	2.8	3.2	1.3
α^b (deg)	0	0	0	0	0	0
β^b (deg)	0	0	0	0	0	0
γ^b (deg)	25	245	135	65	65	205
$P ({}^{14}N)^c$	-3.2	-3.2	3.0	-2.54	-2.40	-2.40
$\eta ({}^{14}\mathrm{N})^c$	-0.63	-0.63	-0.67	0.09	0.09	0.09
α^b (deg)	0	0	0	0	0	0
β^b (deg)	20	20	0	20	20	20
γ^b (deg)	25	245	135	65	65	205

"Units for A (¹⁴N) and P (¹⁴N) values are MHz, which can be converted to 10⁻⁴ cm⁻¹ by dividing the value by 2.997 92. ^bEasyspin Euler angles (α , β , γ) correspond to a rotation about the "gz" axis, a rotation about the new "gy" axis, and a rotation about the new "gz" axis, respectively. ^cPrincipal values of the nuclear quadupole tensor **P** are $e^2 qQ/h(4I(2I-1))[-(1-\eta), -(1+\eta), 2]$, I = 1, $P = e^2 qQ/h$, and η varies between 0 and 1.



Figure 3. X-band ($\nu = 9.671 \text{ GHz}$) HYSCORE (335.0 mT) and orientation-selective three-pulse ESEEM spectra of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ in methanol recorded at 5.0 K. (a,b) Surface and contour plots showing the HYSCORE spectrum. (b) A contour plot showing an overlay of the experimental and simulated HYSCORE spectra. ¹⁴N single- and double-quantum cross-peaks are labeled "s" and "d", respectively. A selection of simulated cross-peaks are labeled for three distal ¹⁴N atoms (N-14a (red), N-14b (purple), and N-34 (green)). Atom numbers correspond to those given in Figure 5. For reference the nitrogen Larmor frequency is $\nu(^{14}N) = 1.03 \text{ MHz}$. (c) Orientation selective three-pulse ESEEM spectra (black) recorded at the indicated field positions along with the simulations (blue) for the distal nitrogens. Spin Hamiltonian parameters determined from the simulation of both the HYSCORE and three-pulse ESEEM spectra are given in Table 2

Computer simulation of the HYSCORE and orientationselective three-pulse ESEEM spectra (Figure 3), assuming the two peaks labeled N-14a and N-14b (Figure 3b) arise from a single nitrogen atom (N-14) in the *N*-methylimidazole ring (see Supporting Information, Figure S4), fails to reproduce the double peaks seen in the experimental spectrum. In contrast, computer simulation of the HYSCORE and orientationselective three-pulse ESEEM spectra with an anisotropic spin Hamiltonian (eq 1) with three inequivalent ¹⁴N nuclei (N-14a (red), N-14b (purple), and N-34 (green)) and the spin Hamiltonian parameters listed in Table 2 produces simulated spectra (Figure 3b,c) that are in excellent agreement with the experimental spectra. The magnetic inequivalence of the two distal nitrogen nuclei (N-14 and N-34) in the *N*-methyl-imidazole rings is also observed in the DFT calculations (see below).⁵⁵ The ¹⁴N hyperfine interactions have an appreciable isotropic component (A_{iso} (N-14a,b; N-34) = 2.61, 3.00, and 1.10 MHz) signifying that a small percentage of the spin

Article



Figure 4. Calculated and experimental (a) g and (b) |A| (⁶³Cu) matrices for $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ calculated with various combinations of functionals and basis sets with the programs ORCA⁴³ and MAG-ReSpect.⁴⁴ The experimental values are depicted as horizontal lines.

density is delocalized onto the distal nitrogens of the coordinated *N*-methylimidazole rings of the macrocycle. The origin of the two ¹⁴N hyperfine couplings, labeled N-14a and N-14b, will be discussed in conjunction with the DFT studies, below.

Mims and Peisach produced a plot of P versus the asymmetry parameter (η) and showed that the experimental ¹⁴N quadrupole parameters could generally be grouped into three regions, namely, A-deprotonated imidazoles; B-imidazoles coordinated to metal ions; and C-protonated imidazoles.⁵⁶ The ¹⁴N quadrupole parameters (Table 2) for the distal nitrogen atoms in the coordinated N-methylimidazole ring in $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ fall just outside of region B, which is clearly not correct, and this is attributed to N-methylation of the distal imidazole nitrogen reducing the asymmetry of the electric field gradient at the nitrogen nucleus to almost zero (η = 0.09). A similar observation has been made for Cu^{II} diethylenetriamine-substituted imidazole complexes (Im: |P| = 1.43 MHz, $\eta = 0.94$; N-MeIm: |P| = 2.06 MHz, $\eta = 0.20$).⁵⁷ This is in contrast to that observed for Cu^{II} centers found in transition metal ion complexes⁵⁷ and metalloproteins,⁵⁸ for example, phenylalanine hydroxylase⁵⁹ and β -amyloid.^{60,61}

A systematic DFT study of square planar Cu^{II} diethylenetriamine imidazole complexes by Ames and Larsen⁵⁵ shows that the magnitude of the distal ¹⁴N hyperfine and quadrupole couplings depend on the orientation of the plane of the imidazole ring with respect to the plane of the $d_{x^2-\nu^2}^2$ orbital. Maximal and minimal ¹⁴N hyperfine and quadrupole couplings were observed when the ring was perpendicular and parallel to the $d_{x^2-y^2}$ orbital, respectively.⁵⁵ In contrast, the lower symmetry (distorted square pyramidal) of the Cu^{II} center in $[Cu^{II}(H_2L^1)$ - $(MeOH)_2$ ⁺ and the relative position of the distal nitrogens in the coordinated N-methylimidazaole rings (constrained by the cyclic peptide) with respect to the Cu^{II} ion yields DFTcalculated ¹⁴N hyperfine and quadrupole couplings (Tables 2 and 4), which are not in agreement with those of the DFT study of square planar Cu^{II} diethylenetriamine imidazole complexes.⁵⁵ Thus, the precise nature of the SOMO, orientation of the N-methylimidazole rings, and methylation of the distal nitrogens will affect the ¹⁴N hyperfine and quadrupole couplings of the ligating and distal nitrogens in the coordinated N-methylimidazole rings.56

DFT Calculation of EPR Spin Hamiltonian Parameters. Calculation of spin Hamiltonian parameters using ab initio or DFT calculations has become quite routine for many transition metal ion complexes and can lead to many insights into their electronic structure.^{62,63} However, a quantitative calculation of the spin Hamiltonian parameters for Cu^{II} complexes is difficult,



Figure 5. Structures and SOMOs for (a,b) $[Cu^{II}(H_2L^1)(MeOH)_2]^+$, (c,d) $[Cu^{II}(H_2L^2)(MeOH)_2]^+$, (e,f) $[Cu^{II}(H_2L^3)(MeOH)_2]^+$, and (g,h) $[Cu^{II}(H_2L^{wa})(MeOH)_2]^+$ showing delocalization of the unpaired electron onto the *N*-methylimidazole, thiazole, oxazole, and oxazoline rings of the [18]azacrown-6 macrocyclic rings. Nitrogen atom numbers given in red.

and the resulting values can vary dramatically from the experimentally determined values.⁶⁴ To explore the role of the functional and basis set in the calculation of the g and A matrices for Cu^{II} complexes of H_3L^{1-3} and H_3L^{wa} , we have undertaken a range of DFT calculations utilizing ORCA 2.6⁴³ and MAG-ReSpect 1.2.⁴⁴ These calculations were performed on fully DFT geometry-optimized structures (G03,⁴⁰ B3LYP/6-31g^{*}) of the respective complexes.

The spin Hamiltonian parameters (g and $A(^{63}Cu)$ matrices, Table 1)²² of the mononuclear Cu^{II} complexes of H₃L¹⁻³ and H₃L^{wa} have been determined previously from computer simulation of the CW EPR spectra using the computer simulation software suites XSophe–Sophe–XeprView^{36,37} (version 1.1.4) and Molecular Sophe^{37,38} (version 2.0.91) in conjunction with the spin Hamiltonian for an isolated paramagnetic center (H_A):

$$H_{A} = \underline{\mathbf{B}} \bullet g \bullet \underline{\mathbf{S}}^{T} + \underline{\mathbf{S}} \bullet A(^{63,65}Cu) \bullet \underline{\mathbf{I}}^{T}$$
$$- g_{n}\beta_{n}\underline{\mathbf{B}} \bullet \underline{\mathbf{I}}^{T}(^{63,65}Cu) + \sum_{i=1}^{3} (\underline{\mathbf{S}} \bullet A(^{14,15}N) \bullet \underline{\mathbf{I}}_{i}^{T}$$
$$- g_{u}\beta_{n}\underline{\mathbf{B}} \bullet \underline{\mathbf{I}}_{i}^{T}(^{14,15}N))$$
(1)

The computed g matrices for $[Cu^{II}(H_2L^1)(MeOH)_2]^+$, derived from DFT calculations with various combinations of functionals and basis sets, are listed in Table 3 and are shown

graphically in Figure 4a. Corresponding Tables and Figures for $[Cu^{II}(H_2L^{2,3,wa})(MeOH)_2]^+$ are given in the Supporting Information (Tables S1–S3 and Figures S1–S3).

The influence of the amount of exact exchange in the functional on the calculated *g* matrices was probed by using pure GGA functionals, functionals with various amounts of exact exchange (i.e., B3LYP (25% HF) vs B1LYP (20% HF)), and modified BXLYP functionals, where X corresponds to the percentage of exact exchange. The functionals PBE,^{65,66} B3PW,^{45,67–69} TPSS,⁷⁰ B3LYP,^{45,46,48} B1LYP,⁷¹ B38LYP, B40LYP, and BHLYP in combination with the basis sets SVP,⁴⁷ TZV,⁴⁸ TZVP,⁴⁸ 6-31g^{*,72} 6-311g^{*,72} IGLO-III,⁷³ IGLO-III,⁷³ EPR-II,⁷⁴ and Wachters⁷⁵ were also used in the calculations.

There was a large variation in predicted g values among the functionals. The general overestimation of the g shift (especially g_z) has been attributed to a combination of too much covalent bonding and the over prediction of the d-orbital splitting, and consequently hybrid functionals like B3LYP (usually with at least 20% Hartree–Fock exchange) are certainly to be preferred.⁷⁶ However, elevated levels of HF exchange are problematic since they also lead to strong spin contamination.⁷⁷

It was found that increasing the amount of exact exchange to at least 38% is mandatory to obtain g_z values in a reasonable range (Figure 4a). The best agreement of g_z and g_y with the experimentally derived values is obtained using the B40LYP functional. Although in most of the cases the g matrices are overestimated by the BHLYP functional (50% HF exchange), this was used for further investigations. BHLYP is a wellestablished functional, and its various dependencies are wellunderstood. Manually changing the amount of HF exchange might have some unforeseen influence on the calculation, and thus the application of an established functional was considered reasonable. The MAG-ReSpect calculated g matrices of $[\,Cu^{II}(H_2L^1)(MeOH)_2\,]^+$ are in best agreement with the experimentally determined values, especially g_z , which is in excellent agreement with the experiment, albeit with a 10-fold increase in computational time.

The EPR-II basis set of Barone is commonly applied in $A(^{63}\text{Cu})$ matrix calculations.⁷⁴ However, for the investigated Cu^{II} complexes of H₃L¹⁻³ and H₃L^{wa}, it was found that the basis set limit is reached at the IGLO-II level (Figure 4b, Supporting Information, Figures S1-S3, Table 3, and Supporting Information, Tables S1-S3). Utilization of the IGLO-III and EPR-II basis sets basically provided identical results to those of IGLO-II. Analyzing the data (comparison of the experimental and DFT-calculated A_i (i = x, y, z) values) it emerges that a basis set without a frozen core region is important as otherwise the A_z values are in good agreement with the experiment, but the A_{x,y} values are greatly overestimated (e.g., TZVP and 6-31g*). Applying the Wachters basis set to Cu^{II} significantly improves the calculation, reproducing the observed trend that A_{xy} is significantly smaller than A_{zy} yet A_z is overestimated by a factor of $\sim 2.^{75}$ This large overestimation is caused by neglecting spin-orbit coupling (SOC) due to Cu^{II}, and the results are substantially improved when SOC contributions are included. The calculated $A(^{63}Cu)$ matrices for the Cu^{II} complexes of H_3L^{1-3} and H_3L^{wa} and their dependence on the applied basis sets is shown in Table 3 and Supporting Information, Tables S1-S3 and are depicted in Figure 4b and Supporting Information, Figures S1-S3. The inability to accurately define the s-electron spin density at the nucleus (and hence $A_{iso}(^{63}Cu)$ produces inaccurate $A(^{63}Cu)$ matrices (Table 3),

Table 3. Calculated and Experimental g and $A(^{63}Cu)$ Values for the Mononuclear Cu^{II} Complex $[Cu^{II}(H_2L^1)(MeOH_2)]^+$

	g ^a values			A(⁶³ Cu) values ^b			
	g _x	<i>gy</i>	g _z	$ A_x $	lA _y l	Az	
experimental values ²²	2.088	2.051	2.278	17	15	153	
functional/basis set							
PBE/SVP	2.027	2.039	2.105	49	63	131	
PBE/6-311g*	2.026	2.036	2.101	61	73	122	
TPSS/6-311g*	2.026	2.035	2.097	61	76	129	
TPSS/6-311g* ^c TZVP ^{d,e}	2.029	2.040	2.104	93	110	111	
B3PW/6-311g*	2.041	2.052	2.148	63	79	161	
B3PW/6-311g* ^c TZVP ^{d,e}	2.046	2.059	2.163	94	111	146	
B3LYP/6-311g* ^c Wachters ^e	2.043	2.055	2.155	3	19	252	
B3LYP/6-311g*	2.040	2.051	2.146	65	81	153	
B1LYP/SVP	2.046	2.059	2.163	47	66	182	
B1LYP/SVP ^c Wachters ^e	2.047	2.061	2.171	67	84	168	
B38LYP/IGLO-II ^c Wachters ^e	2.071	2.084	2.248	10	30	290	
B40LYP/IGLO-II ^c Wachters ^e	2.074	2.088	2.259	9	30	295	
B40LYP/IGLO-III ^c Wachters ^e	2.072	2.085	2.254	11	29	294	
BHLYP/IGLO-II ^c Wachters ^e	2.085	2.100	2.304	10	33	312	
BHLYP/EPR-II ^c Wachters ^e	2.084	2.099	2.303	10	34	312	
BHLYP/IGLO-II ^c TZVP ^d Wachters ^e	2.090	2.100	2.310	10	33	312	
BHLYP/IGLO-II ^c Wachters ^e SOC	2.100	2.080	2.306	13	31	175	
MAG-ReSpect (BHandHLYP/TZVP)	2.093	2.076	2.278	21	5	133	

^{*a*}Calculated with program packages MAG-ReSpect⁴⁴ and ORCA.⁴³ The dependence of the *g* factors on the functional is reflected in the large deviation of the calculated values. The most accurate (sum of squared differences between experiment and calculated) predicted parameters are denoted in bold. ^{*b*}Units, 10^{-4} cm⁻¹. ^{*c*}C, H, N, O. ^{*d*}first coordination sphere. ^{*e*}Cu.

Table 4. Experimental and Calculated (ORCA⁴³) $A({}^{14}N)$ and $P({}^{14}N)$ Values for the Mononuclear Cu^{II} Complex $[Cu^{II}(H_2L^1)(MeOH_2)]^+ a^*$

	atom number ^b									
			N _{distal} -14					N _{distal} -34		
functional/basis sets	$ A_x $	$ A_y $	$ A_z $	Р	η	$ A_x $	$ A_y $	$ A_z $	Р	η
experimental values ^c	2.5	2.5	2.8	-2.54	0.09	1.0	1.0	1.3	-2.4	0.09
	2.9	2.9	3.2	-2.40	0.09					
BHLYP IGLO-II Cu Wachters	1.5044	1.6332	2.1378	-2.753	0.122	0.3185	0.4379	0.7509	-2.73	0.089
B1LYP 6311gs	1.6419	1.7922	2.451	-2.498	0.128	0.2494	0.4052	0.7379	-2.466	0.095
B1LYP 6311gs Cu Wachters	1.5919	1.7403	2.3802	-2.498	0.128	0.2301	0.3883	0.7185	-2.465	0.095
PBE 6311g*	1.6945	1.89	2.6426	-2.517	0.142	0.3005	0.4736	0.7802	-2.467	0.129
PBE 6311g* TZV	1.7284	1.9272	2.6298	-2.527	0.144	0.3802	0.554	0.8657	-2.477	0.132
TPSS 6311g*	1.6222	1.8085	2.5868	-2.503	0.136	0.1918	0.3716	0.6912	-2.456	0.133
TPSS 6311g* TZV	1.6506	1.8387	2.5653	-2.513	0.139	0.2687	0.4471	0.7698	-2.465	0.118
B3PW 6311g*	1.727	1.8839	2.5745	-2.517	0.133	0.2351	0.4091	0.7405	-2.485	0.107
B3PW 6311g* TZV	1.7093	1.8687	2.516	-2.526	0.137	0.2615	0.4402	0.7732	-2.493	0.112
B3LYP 6311g*	1.6915	1.85	2.5417	-2.494	0.13	0.2599	0.421	0.7549	-2.458	0.1
B1LYP SVP	2.451 ^d	2.6075	3.2561	-2.542	0.126	0.3016	0.4958	0.8233	-2.511	0.097
B1LYP 6311g+ Wachters	1.5919	1.7403	2.3802	-2.498	0.128	0.2301	0.3883	0.7185	-2.465	0.095
BHLYP EPRII Cu Wachters	1.5039	1.6262	2.1294	-2.678	0.119	0.2752	0.4032	0.7169	-2.665	0.083

^{*a*}Anisotropic ¹⁴N hyperfine (*A*) and quadrupole (*P*) parameters have units of MHz. ^{*b*}Nitrogen atom numbers are given in red in Figure 5. ^{*c*}There are two sets of parameters for N_{distal} -14, potentially arising from two different conformations of the *N*-methylimidazole ring. ^{*d*}The most accurate (sum of squared differences between experiment and calculated) predicted parameters are denoted in bold.

and consequently results in inaccurate $A(^{14}N)$ hyperfine couplings from ligating nitrogen atoms (Supporting Information, Table S4). However, Lancaster et al. showed that QM/ MM in conjunction with spectroscopy-oriented configuration interaction (SORCI) accurately reproduced the g and 63 Cu and ^{14}N hyperfine matrices for the type zero Cu^{II} center in variants of *Pseudomonas aeruginosa* azurin.⁶³ In contrast, the $A(^{14}N)$ hyperfine couplings from the distal nitrogen atoms in the *N*methylimidazole rings mainly arise from a dipole–dipole interaction. This is confirmed in the calculations of the anisotropic $A(^{14}N)$ hyperfine and $P(^{14}N)$ nuclear quadrupole matrices (Table 4), where there is significantly smaller variation and that all of the results are in reasonable agreement with the experimental values (Tables 2 and 4) obtained from the HYSCORE and three-pulse ESEEM spectra. Spin densities (and the ¹⁴N hyperfine couplings) on the noncoordinated amide and *N*-methylimidazole nitrogen atoms are significantly smaller at least by a factor of 10 smaller) than those for distal nitrogen atoms of the coordinated *N*-methylimidazole ligands (Figure 5). Consequently, a plausible explanation for the observation of two ¹⁴N hyperfine couplings, labeled N-14a and N-14b, is that there are two conformations of the coordinated *N*-methylimidazole ring. Rotation of the ring must be quite small as the difference between the spin Hamiltonian parameters (N-14a vs N-14b) is quite small (Table 2). Confirmation of this would require ¹⁵N NMR studies of the paramagnetic Cu^{II} complex, followed by molecular modeling to find the local minima on the potential energy surface and subsequent DFT studies to optimize the geometries and calculate the spin Hamiltonian properties. This is beyond the scope of the current research.

In agreement with the HYSCORE and ESEEM results, the DFT calculations reveal that the distal nitrogens, N-14 and N-34 in the coordinated N-methylimidazole rings are magnetically inequivalent (Table 4), which is reflected in their different spin densities in the singly occupied molecular orbital (SOMO, Figure 5b). The different ¹⁴N hyperfine couplings result from the N-methylimidazole ring (containing N-34) being twisted out of the equatorial plane containing the Cu^{II} $d_{x^2-y^2}$ orbital and N-34 being oriented between the Cu–ligand bonds (Figure 5a), rather than approximately along the Cu–ligand bond as for N-14. These factors result in reduced isotropic and anisotropic ¹⁴N hyperfine couplings for N-34. The DFT calculations (Table 4) also reproduce the experimental quadrupole parameters (P, η) indicating that indeed methylation of the distal nitrogen reduces the asymmetry parameter to approximately zero.

A comparison of the SOMO values for the Cu^{II} complexes $([Cu^{II}(H_2L^1)(MeOH)_2]^+, [Cu^{II}(H_2L^2)(MeOH)_2]^+, [Cu^{II}(H_2L^3)(MeOH)_2]^+, and [Cu^{II}(H_2L^{wa})(MeOH)_2]^+) (Fig$ ure 5) reveals subtle differences in the amount of spin density on the distal heteroatoms in the rings (N-methylimidazole, oxazole, thiazole, and oxazoline) coordinated to the Cu^{II} ion through the nitrogen atom. For all complexes the distal heteroatoms in the two coordinated heterocyclic rings have inequivalent spin densities $([Cu^{II}(H_2L^1)(MeOH)_2]^+$ - N-14: 3.475×10^{-3} , N-34: -5.37×10^{-4} ; $[Cu^{II}(H_2L^2)(MeOH)_2]^+$ - $O-14:3.631 \times 10^{-3}, O-34:1.98 \times 10^{-4}; [Cu^{II}(H_2L^3)(MeOH_2)]^+$ - S-14:3.990 × 10^{-3} , S-34:3.83 × 10^{-4} ; [Cu^{II}(H₂L^{wa})- $(MeOH)_2$]⁺ - O-14:3.218 × 10⁻³, O-34:4.66 × 10⁻⁴), which originates from the geometric and electronic factors described above for $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ (Figure 5a,c,e,g). Interestingly but not unexpectedly, there is more spin density on the distal sulfur atoms (S-14, S-34) of the coordinated heterocyclic rings as the 3p orbitals are larger than those for N and O (2p orbitals), enabling greater delocalization of the unpaired electron spin. While the spin densities are small, the agreement between the experimental and calculated ¹⁴N hyperfine and nuclear quadrupole splittings of the distal nitrogen in the heterocyclic rings of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ is excellent giving confidence in the spin densities for the distal atoms in the heterocyclic rings of the other complexes.

MCD Spectroscopy. An MCD experiment provides simultaneous information about the ground and excited states of a paramagnetic material.^{78,79} A spectrum can be observed when circularly polarized light is propagated through a sample positioned within a strong magnetic field parallel to the direction of propagation. The MCD spectrum is detected as the absorbance difference of the substance between left and right circularly polarized light.⁸⁰ The general equation that describes the MCD signal is

$$\frac{\Delta \varepsilon}{E} = \frac{\varepsilon_{\rm LCP} - \varepsilon_{\rm RCP}}{E}$$
$$= \gamma \mu_{\rm B} B \left[\left(-\frac{\partial f(E)}{\partial E} \right) \overline{A}_1 + \left(\overline{B}_0 + \frac{\overline{C}_0}{kT} \right) f(E) \right]$$
(2)

where γ is a collection of constants, B = magnetic flux density, k = Boltzmann constant, $\mu_{\rm B} =$ Bohr magneton, f(E) = line shape function, T = temperature, $\varepsilon_{\rm LCP}$ and $\varepsilon_{\rm RCP}$ are the extinction coefficients for left and right circularly polarized light.^{78–82} A_1 , B_0 , and C_0 are characteristic terms that depend on the electronic and geometric structure of the molecule under investigation. Each molecule with degenerate excited states exhibits an A_1 term with a derivative band shape. While the B_0 term has an absorption-like band shape and arises when the applied field causes a mixing of the ground state or an excited state with an intermediate state, the C_0 term is caused by degenerate ground states and is therefore only present in paramagnetic species. It is the only temperature-dependent term, dominating the spectra at low temperatures and identifies paramagnetic species.

Variable-temperature MCD spectra (Figure 6) of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ reveal a strong temperature depend-



Figure 6. Variable-temperature MCD spectra of $[Cu^{II}(H_2L^1)-(MeOH)_2]^+$ measured at 5 T in methanol/glycerol (1:1) frozen solution, $c[Cu^{II}(H_2L^1)(MeOH)_2]^+) = 30$ mM. (inset) VTVH saturation curves at $\lambda = 700$ nm, experimental (black crosses), and simulated (red lines). All curves are overlaying.

ence of the signals consistent with a large C_0 term arising from the paramagnetic Cu^{II} center. The MCD saturation curve of the absorption at 700 nm and its fit (red) is shown in the inset to Figure 6. It was fitted assuming $g_z = 2.283$ and an *xy* polarized transition. This is in agreement with the simulated EPR data (Table 1; $g_z = 2.278$). The MCD spectra of $[Cu^{II}(H_2L^1)-(MeOH)_2]^+$ are in good agreement with the experimental findings from the CW and pulsed EPR, UV–vis, and CD spectroscopic studies,^{22,83} which are characteristic of a square pyramidal Cu^{II} coordination geometry with Cu^{II} bound to the N_{het}–N_{amide}–N_{het} motif.

While the EPR measurements and DFT calculations of the geometry and spin state for the dimer $[Cu_2^{II}(L^1)(\mu$ -OMe)] predict a strongly antiferromagnetically coupled species that is EPR silent at 77 K, the MCD data measured in a methanol/glycerol (1:1) glass (Figure 7) are consistent with a weakly



Figure 7. Variable-temperature MCD spectra of $[Cu_2^{II}(L^1)(\mu-OHCH(CH_2OH)_2)]^+$. (a) Experimental spectra measured at 5 T in methanol/glycerol (1:1) frozen solution, $(c(H_3L^1) = 30 \text{ mM})$. (b) Variable-temperature variable-field curves (black = experimental, red = fit; $\lambda = 660 \text{ nm}$; T = 1.75 K - 50 K, measured at 7 T. Parameters are given in Table 5.

coupled species (nested variable-temperature variable-field (VTVH) curves), and consequently it should be possible to measure an EPR spectrum at 77 K. The possibility that the recorded MCD spectrum arises from an impurity of the monomeric Cu^{II} complex $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ in the sample can be excluded for several reasons. The signal that was recorded for the dimeric complex is strong, and the delta absorption values (ΔA) are similar to those of the monomeric species, while a signal arising from an impurity is expected to have considerably smaller Δ absorption values. Furthermore, the peak positions and general appearance of the MCD spectra (Figures 6 and 7a) of the mono- and the dinuclear species are significantly different. The mononuclear complex $[Cu^{II}(H_2L^1) (MeOH)_2$ ⁺ has a negative Δ absorption band at 700 nm and a positive Δ absorption at 940 nm, whereas the dinuclear Cu^{II} complex of H_3L^1 exhibits two negative Δ absorption bands at 550 and 660 nm. Moreover, the VTVH curves of the dinuclear Cu^{II} complex, measured at 660 nm, are nested, which is a strong indication for coupled species (Figure 7b).

From the experimental VTVH curves of the dinuclear Cu^{II} complex of H_3L^1 , the *g* matrix, exchange-coupling constant *J*, and the axial (D) and rhombic (E) zero-field splitting values can be derived from a fit of the nested VTVH curves. The VTVH data was analyzed using the formalism developed by

Neese and Solomon, eq 3.⁷⁸ For a particular coupled spin system S_1 and S_2 , the MCD magnetization curves are calculated as

$$\frac{\Delta A}{E} = -\frac{1}{4\pi S} \int_0^{\pi} \int_0^{2\pi} \sum_i N_i \left(l \langle S_{xli} \rangle M_{yz1}^{\text{eff}} + m \langle S_{yli} \rangle M_{xz1}^{\text{eff}} \right)$$
$$+ n \langle S_{zli} \rangle M_{yx1}^{\text{eff}} + l \langle S_{x2i} \rangle M_{yz2}^{\text{eff}} + m \langle S_{y2i} \rangle M_{xz2}^{\text{eff}}$$
$$+ n \langle S_{z2i} \rangle M_{yz2}^{\text{eff}} \rangle \sin \theta d\theta d\phi + B_0 B$$
(3)

where N_i are the Boltzmann populations of the energy levels; $\langle S_{xli} \rangle$ is the expectation value of the spin operator S_x for spin 1 in level *i*; *l*, *m*, *n* specifies the direction of the magnetic field with respect to a fixed molecular axis system; and the integration is carried out numerically over all orientations. The M_{xy}^{eff} , M_{xz}^{eff} , M_{yz}^{eff} are the effective transition dipole moment products for transition allowed in *x* and *y* polarization, one expects $M_{xy}^{\text{eff}} \gg M_{xz}^{\text{eff}} \approx M_{yz}^{\text{eff}}$. The last term describes the field-dependent B_0 term of eq 2. The population factors N_i and the spin expectation values are calculated from the energies and wave functions, respectively, of the spin Hamiltonian for the coupled system

$$H = \sum_{i=1}^{2} H_{Ai} + H_{int}$$
$$H_{int} = -2J_{iso}\underline{S}_{1} \bullet \underline{S}_{2} + \underline{S}_{1} \bullet D \bullet \underline{S}_{2}$$
(4)

where H_A is given in eq 1. The best fit of the experimental data is shown in Figure 7b. For coupled spin systems the D matrix (quantified by zero-field splitting D, E values) is nonzero due to a combination of anisotropic exchange (J) and dipole-dipole coupling terms. Without additional structural information about the metal-metal distance and orientation of principal axes, it is difficult to separate these terms. We therefore fitted the dimer VTVH data (Figure 7b) with an effective S = 1 spin Hamiltonian resulting in the parameters given in Table 5. The quality of the fit indicates that the Cu^{II} ions are not strongly antiferromagnetically coupled as population of the higher $S_{\text{Tot}} =$ 1 spin state would be observed in the VTVH data. Thus, the Cu^{II} ions are either very weakly exchange coupled or strongly

Table 5. Experimentally Determined Parameters from MCD and EPR for the Coupled Dimer $[Cu_2^{II}(L^1)(\mu$ -OHCH $(CH_2OH)_2)]^+$ Complex in a Methanol/Glycerol (1:1) Solvent Mixture, Treating It as an Effective S = 1 spin System (see text)^{*a,b*}

parameter ^a	value
g_x^b	2.121
g_y^b	2.155
g_z^b	2.130
$ A_x ({}^{63}Cu)^b$	$20.3 \times 10^{-4} \text{ cm}^{-1}$
$ A_{y} ({}^{63}Cu)^{b}$	$21.6 \times 10^{-4} \text{ cm}^{-1}$
$ A_z $ (⁶³ Cu) ^b	$150 \times 10^{-4} \text{ cm}^{-1}$
D	0.0185 cm^{-1}
E/IDI	0.220
B_0	-0.0011 T^{-1}

^{*a*}The parameters simultaneously fit the MCD (Figure 7b) and the EPR (Figure 8b) except for $|A_i|$ (⁶³Cu) (EPR only) and B_0 (MCD only). ^{*b*}The *g*- and *A*-values refer to an effective *S* = 1 spin Hamtionian (eq 4) of a Cu^{II} dimer system.



Figure 8. X-Band EPR spectra of $[Cu_2^{II}(L^1)(\mu$ -OHCH $(CH_2OH)_2)]^+$. (a) Experimental EPR spectrum of $[Cu_2^{II}(L^1)(\mu$ -OHCH $(CH_2OH)_2)]^+$ in a methanol/glycerol (1:1) frozen solution, $c(H_3L^1) = 1.5$ mM, $\nu = 9.434401$ GHz, T = 50 K. (b) Computer simulation of (a). (c–e) Energy level diagrams showing the allowed EPR transitions along the *z*, *x*, and *y* principal directions, respectively. Spin Hamiltonian parameters are given in Table 5

ferromagnetically coupled. The only experimental difference between the MCD and EPR-silent experiment of the dimer system as described above, was the presence of 50% glycerol, to generate an optically clear glass. Repeating the EPR measurements under these conditions produced the EPR spectrum shown in Figure 8, which arises from the coupled dinuclear Cu^{II} complex. When the temperature is decreased to 10 K, saturation of the EPR transitions is observed, which is consistent with a small exchange coupling (Orbach relaxation⁸⁴), predicted by the MCD measurements.

The experimental EPR spectrum could be simulated (Figure 8b) with an S = 1 spin Hamiltonian (eq 4) and the spin Hamiltonian parameters given in Table 5. The corresponding energy level diagrams and transitions along the z, x, and yprincipal directions are shown in Figure 8c-e. A mass spectrum of the dinuclear Cu^{II} complex in methanol/glycerol (1:1) reveals peaks (Supporting Information, Figure S5a,c) at m/z =825.203 58 and 861.054 22. Calculated spectra for $[Cu_2^{II}(L^1) (OCH(CH_2OH)_2)MeOH(H_2O)]^+$ (m/z = 861.28658) and $[Cu_2^{II}(HL^1)(OCH(CH_2OH)_2)MeOH]^+$ (m/z = 825.26545) (Supporting Information, Figure S5b,d) are in excellent agreement, particularly the isotope distributions (Supporting Information, Figure S5). Assuming glycerol bridges the two Cu^{II} ions ([$Cu_2^{II}(L^1)(\mu$ -OHCH($CH_2OH)_2$)]⁺), which leads to reduced exchange coupling, we performed a geometry optimization of the dinuclear complex utilizing DFT methods (B3LYP/6-31g* (C, H, N, O)/TZVP(Cu); Figure 9). This resulted in a calculated exchange coupling constant J_{iso} (-18 cm⁻¹) that was significantly smaller than that for the methoxide-bridged dinuclear complex ($J_{iso} = -147 \text{ cm}^{-1}$). While the calculation is consistent with the experimental conclusion that the $|J_{iso}|$ value is small, and confirms that glycerol binding modifies the exchange coupling between the Cu^{II} ions and produces different spectroscopic signatures in both the MCD and EPR spectra, there are a number of uncertainties in the orientation of the bridging glycerol and the possibility of additional solvent molecule(s).



Figure 9. DFT-calculated structure (Gaussian03,⁴⁰ B3LYP/6-31g*/ TZVP) of the glycerol-bridged dinuclear Cu^{II} complex $[Cu_2^{II}(L^1)(\mu-OHCH(CH_2OH)_2)]^+$.

CONCLUSIONS

High-resolution EPR spectroscopy in conjunction with DFT calculations provides a powerful approach for the geometric and electronic structural characterization of paramagnetic species. Herein we utilized orientation-selective pulsed ENDOR, three-pulse ESEEM, and HYSCORE in conjunction with DFT calculations and the published CW EPR data to determine the geometric and electronic structure of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$. The electronic structures of the other $[Cu^{II}(H_2L^{2-3,wa})(MeOH)_2]^+$ complexes were determined by DFT calculations and utilized the CW EPR data to test the validity of the optimized structure. The geometric structures of $[Cu^{II}(H_2L^{2-3,wa})(MeOH)_2]^+$ were similar to that of $[Cu^{II}(H_2L^{2-3,wa})(MeOH)_2]^+$ involved the third oxazoline nitrogen.

Orientation-selective ENDOR measurements of $[Cu^{II}(H_2L^1)(MeOH)_2]^+$ yielded the ligating nitrogen hyperfine and nuclear quadrupole coupling constants (Table 2), which showed that they were magnetically inequivalent, a consequence of the steric constraints of the ligand and the differing nitrogens (amide vs imidazole). The ¹⁴N_{het} and ¹⁴N_{amide} quadrupole parameters (Table 2) are in agreement with those in the literature for deprotonated imidazole^{58,59} and peptide amide nitrogens.^{54,85} These results are consistent with Cu^{II} being coordinated to a $N_{het}-N_{amide}-N_{het}$ binding site in H_3L^1 , as suggested from the CW EPR studies.²²

HYSCORE and orientation-selective three-pulse ESEEM measurements on this complex also revealed that the distal nitrogens (N-14 and N-34) in the N-methylimidazole rings coordinated to the Cu^{II} ion were also magnetically inequivalent and that there were two populations (conformations, N-14a and N-14b) of the N-methylimidazole ring containing N-14. The relatively large magnetic inequivalence between N-14 and N-34 is a consequence of the N-methylimidazole ring (containing N-34) being twisted out of the equatorial plane containing the $Cu^{II} d_{x^2-y^2}$ orbital and being oriented between the Cu-ligand bonds (Figure 5a), rather than approximately along the Cu-ligand as for N-14. These factors result in reduced isotropic and anisotropic ¹⁴N hyperfine couplings for N-34, which were confirmed with DFT calculations (Table 4). Similar observations were also observed from the spin densities calculated by DFT for the other complexes ($[Cu^{II}(H_2L^2) (MeOH)_2$]⁺, $[Cu^{II}(H_2L^3)(MeOH)_2]^+$ and $[Cu^{II}(H_2L^{wa})_ (MeOH)_2$ ⁺). While the spin densities on the N_{het}-N_{amide}- N_{het} binding site within $[Cu^{II}(H_2L^{wa})(MeOH)_2]^+$ were similar to the other complexes, there was additional spin density on the third coordinated oxazoline nitrogen. The experimentally determined η parameters (0.09) for the distal nitrogen atoms (Table 2) are consistent with *N*-methylation of the imidazole rings.

A systematic study of functionals and basis sets found that the experimental g, $A(^{63}Cu)$ spin Hamiltonian parameters could be reasonably reproduced with MAG-ReSpect⁴⁴ or using ORCA⁴³ with the BHLYP functional in combination with the basis sets 6-31g* (C, H, N, S, O), TZVP (first coordination sphere), and Wachters (Cu). Importantly, spin–orbit coupling and the incorporation of ~38% Hartree–Fock exchange were critical for the accurate prediction of the spin Hamiltonian parameters (g, $A(^{63}Cu)$). Calculation of the distal heterocyclic nitrogen hyperfine and nuclear quadruple couplings were more accurate as the isotropic hyperfine coupling is significantly smaller, and consequently the proportion of Hartree–Fock exchange included in the calculation has only a small effect.

At high base concentrations all macrocycles (H_3L^{1-3}) show the formation of dinuclear methanol (methoxide)-bridged Cu^{II} complexes.²² MCD spectroscopy in conjunction with CW EPR and DFT studies have shown that this bridging ligand is labile and that it can be replaced by glycerol. Upon ligand replacement of the bridging ligand with glycerol, there is a dramatic decrease in the antiferromagnetic exchange coupling between the two Cu^{II} centers. Thus, with an appropriate choice of solvent the dinuclear Cu^{II} complexes of $\dot{H_{3}L^{1-3}}$ maybe able to hydrolyze CO2 and phospho mono- and diesters as has been shown for the corresponding dinuclear Cu^{II} complexes of the cyclic pseudo octapeptides.^{27,28} Indeed, preliminary experiments involving the dinuclear Cu^{II} complex with H₃L¹ show that it is capable of hydrolyzing BDNPP (bis(2,4dinitrophenyl)phosphate). In contrast, the increased flexibility of the oxazoline versus N-methylimidizale, oxazole, and thiazole rings enables H₃L^{wa} to form a mononuclear Cu^{II} center in which the three imidazole and one amide nitrogen atoms are ligated to the Cu^{II} center, which explains the lack of dinuclear Cu^{II} complexes in the mass spectra.²² Thus, Cu^{II} ion transport, homeostasis, and catalysis are possible biological functions for $H_{3}L^{wa}$ in L. bistratum, where high Cu^{II} concentrations are present.

ASSOCIATED CONTENT

Supporting Information

Tables of DFT-calculated spin Hamiltonian parameters for $[Cu(H_2L^{2,3,wa})(MeOH)_2]^+$, corresponding plots of spin Hamiltonian parameters versus functional, HYSCORE and three-pulse ESEEM simulations, and mass spectra for the dinuclear Cu^{II} complex. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*E-mail: Graeme.Hanson@cai.uq.edu.au. Phone: +61-7-3365-3242. Fax: +61-7-3365-3833.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We would like to thank The University of Queensland and the Ruprecht-Karls-Universität Heidelberg for financial support, Dr. B. Martin for critically reading the manuscript, and Miss N. Mehrkens for measuring the mass spectra and performing the preliminary phosphatase hydrolysis experiments. J.H. thanks the Australian Research Council for an award of a Future Fellowship (FT120100421). We would also like to thank the anonymous referee for encouraging us to reexamine the analysis of the HYSCORE and three-pulse ESEEM data.

REFERENCES

- (1) Kott, P. Mem. Queensl. Mus. 1985, 23, 1-440.
- (2) Kott, P. Mem. Queensl. Mus. 1990, 29, 1–266.
- (3) Kott, P. Mem. Queensl. Mus. 1990, 29, 267-298.
- (4) Kott, P. Mem. Queensl. Mus. 1992, 32, 375-620.
- (5) Kott, P. Mem. Queensl. Mus. 1992, 32, 621-655.
- (6) Kott, P. Mem. Queensl. Mus. 2001, 47, 1-407.

(7) van den Brenk, A. L. Ph.D. Thesis, University of Queensland, 1994.

(8) Wright, S. H. R, A.; Feldmann, J.; Krupp, E.; Jaspars, M. Marine Metabolites and Metal Ion Chelation. In *Handbook of Marine Products;* Fattorusso, E., Gerwick, W. H., Taglialatela-Scafati, O., Eds.; Springer-Verlag: Berlin, 2012; pp 861–892.

(9) Davidson, B. S. Chem. Rev. 1993, 93, 1771-1791.

(10) Haberhauer, G.; Rominger, F. Eur. J. Org. Chem. 2003, 3209-3218.

(11) Comba, P.; Dovalil, N.; Gahan, L. R.; Hanson, G. R.; Westphal, M. Dalton Trans. **2014**, *43*, 1935–1956.

(12) Withers, N.; V, W.; Lewin, R. A. *Phycologia* **1978**, *17*, 167–171. (13) Kühl, M.; Larkum, A. W. D. In *Cellular Origin and Life in Extreme Habitats;* Seckbach, J., Ed.; Kluwer Academic Publishers: Dordrecht, 2002; Vol. 3: Symbiosis, Mechanisms and Model Systems, pp 273–290.

(14) Shimada, A.; Y, N.; Kanai, S.; Lewin, R. A.; Maruyama, T. *Phycologia* **2003**, *42*, 193–197.

(15) Schmidt, E. W.; Nelson, J. T.; Rasko, D. A.; Sudek, S.; Eisen, J. A.; Haygood, M. G.; Ravel, J. Proc. Natl. Acad. Sci. U. S. A. 2005, 102, 7315–7320.

(16) Long, P. F.; Dunlap, W. C.; Battershill, C. N.; Jaspars, M. ChemBioChem 2005, 6, 1760–1765.

(17) van den Brenk, A. L.; Fairlie, D. P.; Hanson, G. R.; Gahan, L. R.; Hawkins, C. J.; Jones, A. *Inorg. Chem.* **1994**, *33*, 2280–2289.

(18) van den Brenk, A. L.; Byriel, K. A.; Fairlie, D. P.; Gahan, L. R.; Hanson, G. R.; Hawkins, C. J.; Jones, A.; Kennard, C. H. L.; Moubaraki, B.; Murray, K. S. *Inorg. Chem.* **1994**, *33*, 3549–3557.

(19) van den Brenk, A. L.; Tyndall, J. D. A.; Cusack, R. M.; Jones, A.; Fairlie, D. P.; Gahan, L. R.; Hanson, G. R. *J. Inorg. Biochem.* **2004**, *98*, 1857–1866.

(20) Comba, P.; Cusack, R.; Fairlie, D. P.; Gahan, L. R.; Hanson, G. R.; Kazmaier, U.; Ramlow, A. *Inorg. Chem.* **1998**, *37*, 6721–6727.

(21) Bernhardt, P. V.; Comba, P.; Fairlie, D. P.; Gahan, L. A.; Hanson, G. R.; Lötzbeyer, L. Chem.—Eur. J. **2002**, *8*, 1527–1536.

(22) Comba, P.; Gahan, L. R.; Haberhauer, G.; Hanson, G. R.; Noble, C. J.; Seibold, B.; van den Brenk, A. L. *Chem.—Eur. J.* **2008**, *14*, 4393–4403.

(23) Comba, P.; Dovalil, N.; Haberhauer, G.; Hanson, G. R.; Karod, Y.; Taura, T. J. Biol. Inorg. Chem. 2010, 15, 1129–1135.

(24) Comba, P.; Dovalil, N.; Hanson, G. R.; Linti, G. Inorg. Chem. 2011, 50, 5165–5174.

(25) Comba, P.; Dovalil, N.; Gahan, L. R.; Haberhauer, G.; Hanson, G. R.; Noble, C. J.; Seibold, B.; Vadivelu, P. *Chem.—Eur. J.* **2012**, *18*,

2578–2590.

(26) Comba, P.; Dovalil, N.; Haberhauer, G.; Kowski, K.; Mehrkens, N.; Westphal, M. Z. Allg. Anorg. Chem. (Special Issue Bioinorganic Chemistry) **2013**, 639, 1395–1400.

(27) Comba, P.; Gahan, L. R.; Hanson, G. R.; Maeder, M.; Westphal, M. Dalton Trans. 2014, 43, 3144–3152.

(28) Comba, P.; Gahan, L. R.; Hanson, G. R.; Westphal, M. Chem. Commun. 2012, 9364–9366.

(29) Stanwell, C.; Gescher, A.; Watters, D. Biochem. Pharmacol. 1993, 45, 1753–1761.

(30) Hambley, T. W.; Hawkins, C. J.; Lavin, M. F.; van den Brenk, A.; Watters, D. J. *Tetrahedron* **1992**, *48*, 341–348.

(31) Haberhauer, G.; Drosdow, E.; Oeser, T.; Rominger, F. *Tetrahedron* **2008**, *64*, 1853–1859.

(32) Wipf, P.; Venkatraman, S.; Miller, C. P.; Geib, S. J. Angew. Chem. **1994**, 106, 1554–1556.

(33) Wipf, P.; M, C. P. J. Am. Chem. Soc. 1992, 114, 10975–10977.
(34) Schweiger, A.; Jeschke, G. Principles of Pulse Electron

Paramagnetic Resonance; Oxford University Press: Oxford, U.K., 2001. (35) Hofer, P.; Grupp, A.; Nebenfuhr, H.; Mehring, M. Chem. Phys. Lett. **1986**, 132, 279–282.

(36) Hanson, G. R.; Gates, K. E.; Noble, C. J.; Griffin, M.; Mitchell, A.; Benson, S. J. Inorg. Biochem. 2004, 98, 903–316.

(37) Hanson, G. R.; Noble, C. J.; Benson, S. XSophe - Sophe - XeprView and Molecular Sophe Computer simulation software suites for the analysis of continuous wave and pulsed EPR and ENDOR spectra. In *EPR of Free Radicals in Solids: Trends in Methods and Applications;* Lund, A., Shiotani, M., Eds.; Springer: Heidelberg, 2013; p 223.

(38) Hanson, G. R.; Noble, C. J.; Benson, S., Molecular Sophe, An Integrated Approach to the Structural Characterization of Metalloproteins - The Next Generation of Computer Simulation Software. In *High Resolution EPR: Applications to Metalloenzymes and Metals in Medicine;* Hanson, G. R., Berliner, L. J., Eds.; Springer: New York, **2009**; Vol. 28, pp 105–174.

(39) Stoll, S.; Schweiger, A. J. Magn. Reson. 2006, 178, 42-55.

(40) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. Gaussian 03, Revision B.03; Gaussian, Inc.: Wallingford, CT, 2003.

(41) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. *Gaussian 09*, Revision A.02; Gaussian, Inc.: Wallingford, CT, 2009.

(42) Furche, F.; Ahlrichs, R.; Hättig, C.; Klopper, W.; Sierka, M.; Weigend, F. WIREs Comput. Mol. Sci. 2014, 4, 91–100.

(43) Neese, F. Comput. Mol. Sci. 2012, 2, 73-78.

(44) Malkin, V. G.; Malkina, O. L.; Reviakine, R.; Arbuznikov, A. V.; Kaupp, M.; Schimmelpfennig, B.; Malkin, I.; Repiský, M.; Komorovský, S.; Hrobárik, P.; Malkin, E.; Helgaker, T.; Ruud, R. *ReSpect program*, ver 2.2, 2004.

(45) Becke, A. D. Phys. Rev. A 1988, 38, 3098-3100.

(46) Lee, C.; Yang, W.; Parr, R. G. Phys. Rev. B 1988, 37, 785-789.

(47) Schäfer, A.; Horn, H.; Ahlrichs, R. J. Chem. Phys. 1992, 97, 2571-2577.

(48) Schäfer, A.; Huber, C.; Ahlrichs, R. J. Chem. Phys. 1994, 100, 5829-5835.

(49) Antholine, W. E.; Bennett, B.; Hanson, G. R. Copper Coordination Environments. In *Multifrequency Electron Paramagnetic Resonance;* Misra, S. K., Ed.; Wiley-VCH: Verlag, GmbH, 2011; pp 647–718.

(50) Froncisz, W.; Hyde, J. S. J. Chem. Phys. 1980, 73, 3123-3131.

(51) Hyde, J. S.; Froncisz, W. Annu. Rev. Biophys. Bioeng. 1982, 11, 391-417.

(52) Pilbrow, J. R. J. Magn. Reson. 1984, 58, 186-203.

(53) Pogni, R.; Dellalunga, G.; Basosi, R. J. Am. Chem. Soc. 1993, 115, 1546–1550.

(54) Edmonds, D. T. Phys. Rep. 1977, 29, 233-290.

(55) Ames, W. M.; Larsen, S. C. Phys. Chem. Chem. Phys. 2009, 11, 8266-8274.

(56) Mims, W. B.; Peisach, J. ESEEM and LEFE of Metalloproteins and Model Complexes. In *Advanced EPR Applications in Biology and Biochemistry*; Hoff, A. J., Ed.; Elsevier: Amsterdam, 1989; pp 1–57.

(57) Jiang, F.; McCracken, J.; Peisach, J. J. Am. Chem. Soc. 1990, 112, 9035-9044.

(58) Deligiannakis, Y.; Louloudi, M.; Hadjiliadis, N. Coord. Chem. Rev. 2000, 204, 1-112.

(59) McCracken, J.; Pember, S.; Benkovic, S. J.; Villafranca, J. J.; Miller, R. J. J. Am. Chem. Soc. **1988**, 110, 1069–1074.

(60) Drew, S. C.; Noble, C. J.; Masters, C. L.; Hanson, G. R.; Barnham, K. J. J. Am. Chem. Soc. 2009, 131, 1195–1207.

(61) Drew, S. C.; Barnham, K. J. Acc. Chem. Res. 2011, 44, 1146–1155.

(62) De Visser, S. P.; Quesne, M. G.; Martin, B.; Comba, P.; Ryde, U. Chem. Commun. 2014, 50, 262–282.

(63) Lancaster, K. M.; Zaballa, M.-E.; Sproules, S.; Sundararajan, M.; DeBeer, S.; Richards, J. H.; Vila, A. J.; Neese, F.; Gray, H. B. *J. Am. Chem. Soc.* **2012**, *134*, 8241–8253.

(64) Atanasov, M.; Comba, P.; Martin, B.; Müller, V.; Rajaraman, G.; Rohwer, H.; Wunderlich, S. *J. Comput. Chem.* **2006**, *27*, 1263–1277. (65) Perdew, J. P.; Burke, K.; Ernzerhof, M. *Phys. Rev. Lett.* **1996**, *77*, 3865–3868.

(66) Perdew, J. P.; Burke, K.; Ernzerhof, M. Phys. Rev. Lett. 1998, 80, 891.

(67) Perdew, J. P.; Chevary, J. A.; Vosko, S. H.; Jackson, K. A.; Pederson, M. R.; Singh, D. J.; Fiolhais, C. *Phys. Rev. B* **1992**, *46*, 6671–6687.

(68) Perdew, J. P.; Chevary, J. A.; Vosko, S. H.; Jackson, K. A.; Pederson, M. R.; Singh, D. J.; Fiolhais, C. *Phys. Rev. B* **1993**, *48*, 4978.

(69) Perdew, J. P.; Burke, K.; Wang, Y. Phys. Rev. B 1996, 54, 16533–16539.

(70) Tao, J. M.; Perdew, J. P.; Staroverov, V. N.; Scuseria, G. E. Phys. Rev. Lett. **2003**, 91, 146401–1–146401–4.

(71) Adamo, C.; Barone, V. Chem. Phys. Lett. 1997, 274, 242–250.
(72) Hehre, W. J.; Ditchfield, R.; Pople, J. A. J. Chem. Phys. 1972, 56, 2257–2261.

(73) Kutzelnigg, W.; Fleischer, U.; Schindler, M. The IGLO-Method: Ab-initio Calculation and Interpretation of NMR Chemical Shifts and Magnetic Susceptibilities. In *NMR Basic Principles and Progress;* Diehl, P., Fluck, E., Kosfeld, R., Ed.; Springer-Verlag: Berlin, Germany, 1991; Vol. 23, pp 165–262.

(74) Barone, V. Structure, Magnetic Properties and Reactivities of Open-Shell Species From Density Functional and Self-Consistent Hybrid Methods. In *Recent Advances in Density Functional Methods* (*Part I*); Chong, D. P., Ed.; World Scientific: Singapore, 1996; Vol. 1, pp 287–334.

(75) Watchers, A. J. H. J. Chem. Phys. 1970, 52, 1033-1036.

(76) Neese, F. J. Chem. Phys. 2001, 115, 11080-11096.

(77) Kaupp, M.; Reviakine, R.; Malkina, O. L.; Arbuznikov, A.; Schimmelpfennig, B.; Malkin, V. G. J. Comput. Chem. 2002, 23, 794–803.

(78) Neese, F.; Solomon, E. I. Inorg. Chem. 1999, 38, 1847-1865.

(79) Osborne, G. A.; Stephens, P. J. J. Chem. Phys. 1972, 56, 609-618.

(80) Piepho, S. B.; Schatz, P. N. Group Theory in Spectroscopy with Applications to Magnetic Circular Dichroism; John Wiley and Sons: New York, 1983.

(81) Stephens, P. J. J. Chem. Phys. 1970, 52, 3489-3516.

(82) Stephens, P. J. Chem. Phys. Lett. 1968, 2, 241-244.

(83) Seibold, B. Kupfer(II)-Koordinationschemie zyklischer [24] Azakrone-8- und [18]Azakrone-6-Peptide Ruprecht Karls Universitat 2008. http://www.ub.uni-heidelberg.de/archiv/8637.

(84) Pilbrow, J. R. Transition Ion Electron Paramagnetic Resonance; Clarendon Press: Oxford, 1990.

(85) Grimaldi, S.; MacMillan, F.; Ostermann, T.; Ludwig, B.; Michel, H. Biochemistry (Easton) **2001**, 40, 1037–1043.